

# OPTIMIZATIONS OF DETECTION AND QUANTIFICATION OF GMOS

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# Introduction

National reference laboratories for genetically modified organisms (GMOs) and plant pathogens

Partner in two European Union Reference Laboratories (for viruses and phytoplasmas, and for bacteria)

ISO 17025 since 2003, flexible scope since 2006

Development and validation or verification of methods

Routine analysis (plant pathogens, GMOs: food, feed, seed, plants)

Designated as national metrological institute - Holder of National Standard in the Field of Amount of Substance/Bioanalysis of Nucleic Acids/GMOs and Microorganisms

# Genetically Modified Organisms

...are by the EU legal definition<sup>1</sup>: organisms, with the exception of humans, in which the genetic material has been altered through the use of biotechnological methods, in a way that does not occur naturally by mating and/or natural recombination.



<sup>1</sup> Directive 2001/18/EC of the European Parliament and the Council of 12 March 2001 on the deliberate release in to the environment of genetically modified organisms and repealing Council Directive 90/220/EC

# GMOs in the European Union

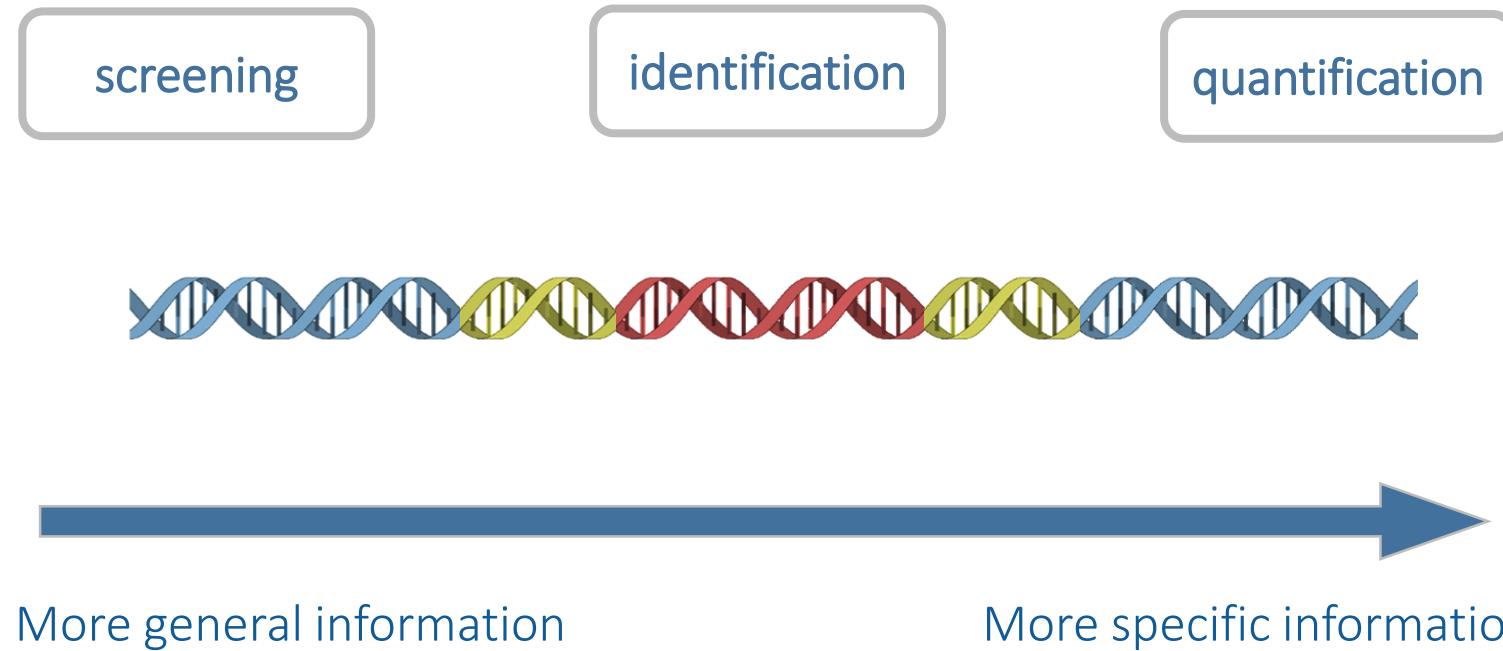
Regulation No. 1829/2003

Labelling "...shall not apply to foods/feed containing material which contains, consists of or is produced from GMOs in a proportion **no higher than 0.9 per cent** of the food ingredients considered individually or food consisting of a single ingredient..."

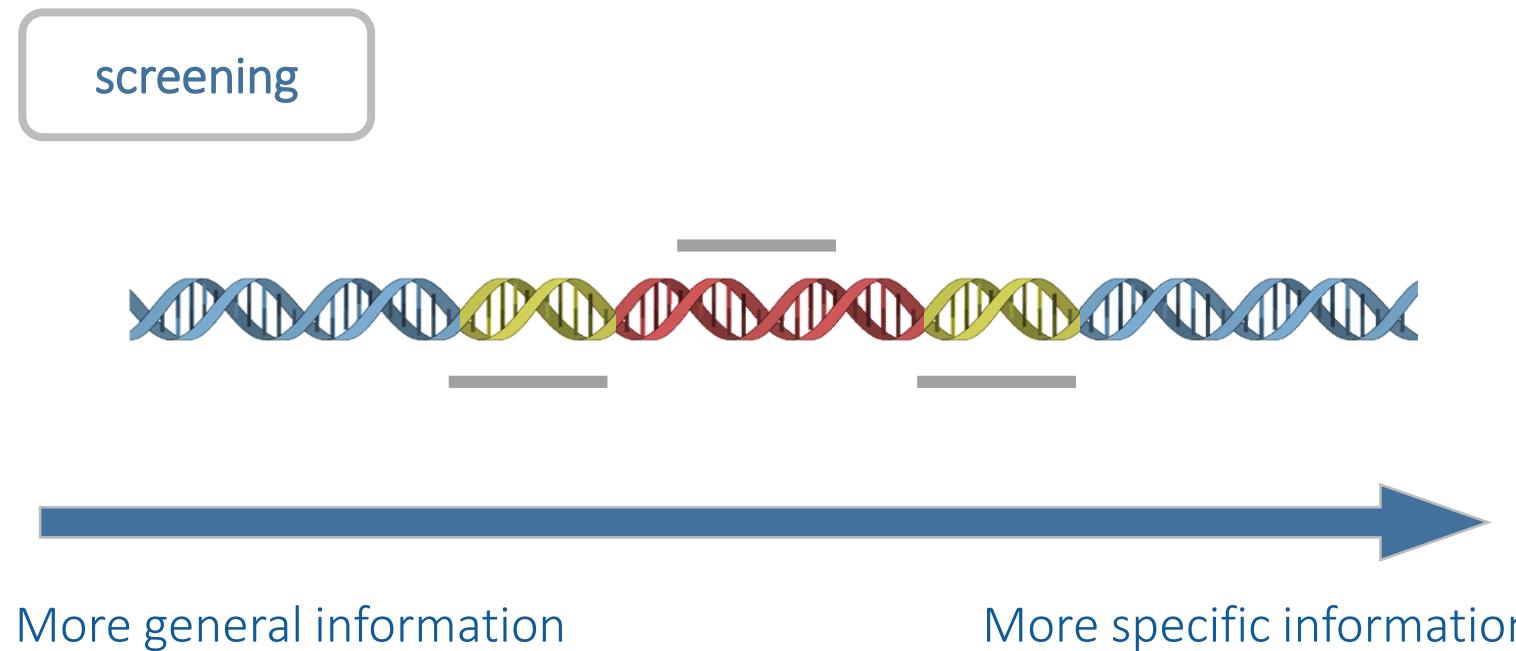
Regulation No. 619/2011 (so called LLP regulation)

"...it is appropriate to set as a Minimum Required Performance Limit (MRPL) the lowest level of GM material which is considered by the EU-RL for the validation of quantitative methods. This level corresponds to **0.1% related to mass fraction** of GM material in feed and is the lowest level where results are satisfactorily reproducible between official laboratories ..."

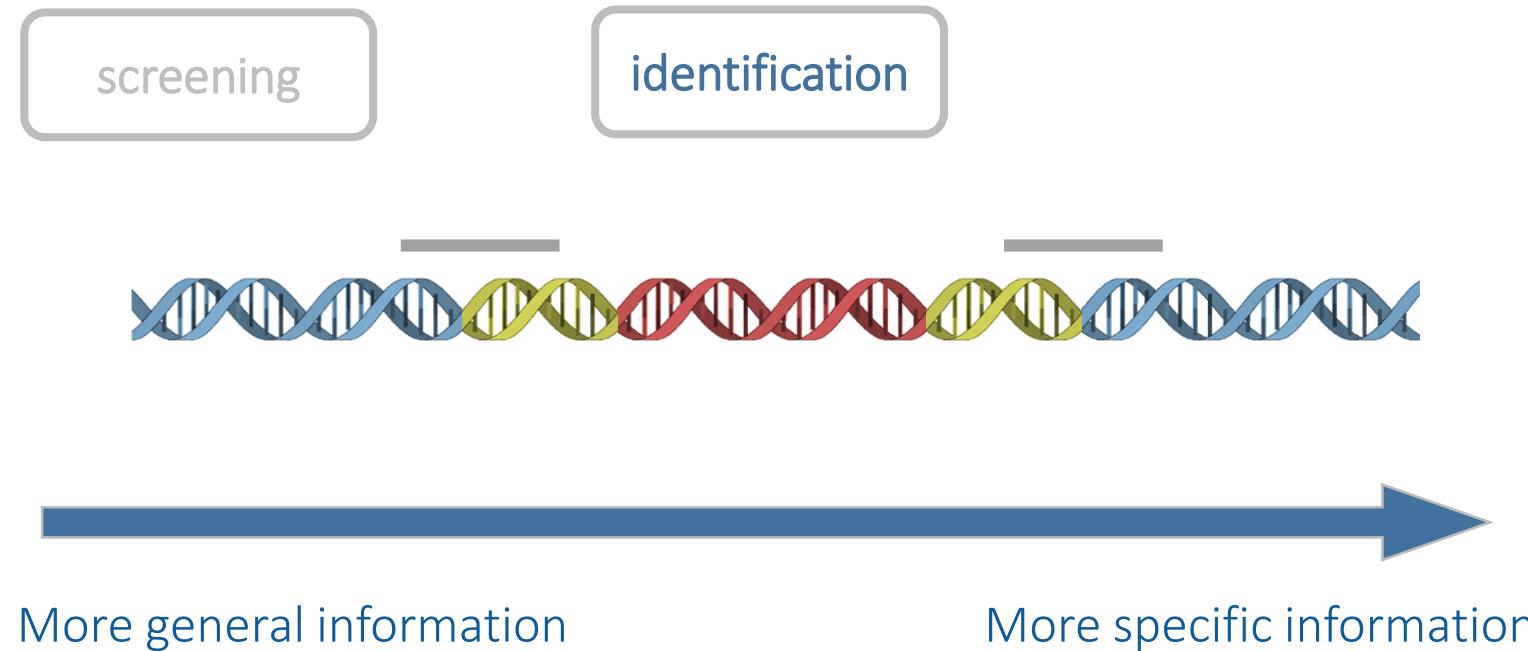
# Three partite PCR-based testing scheme



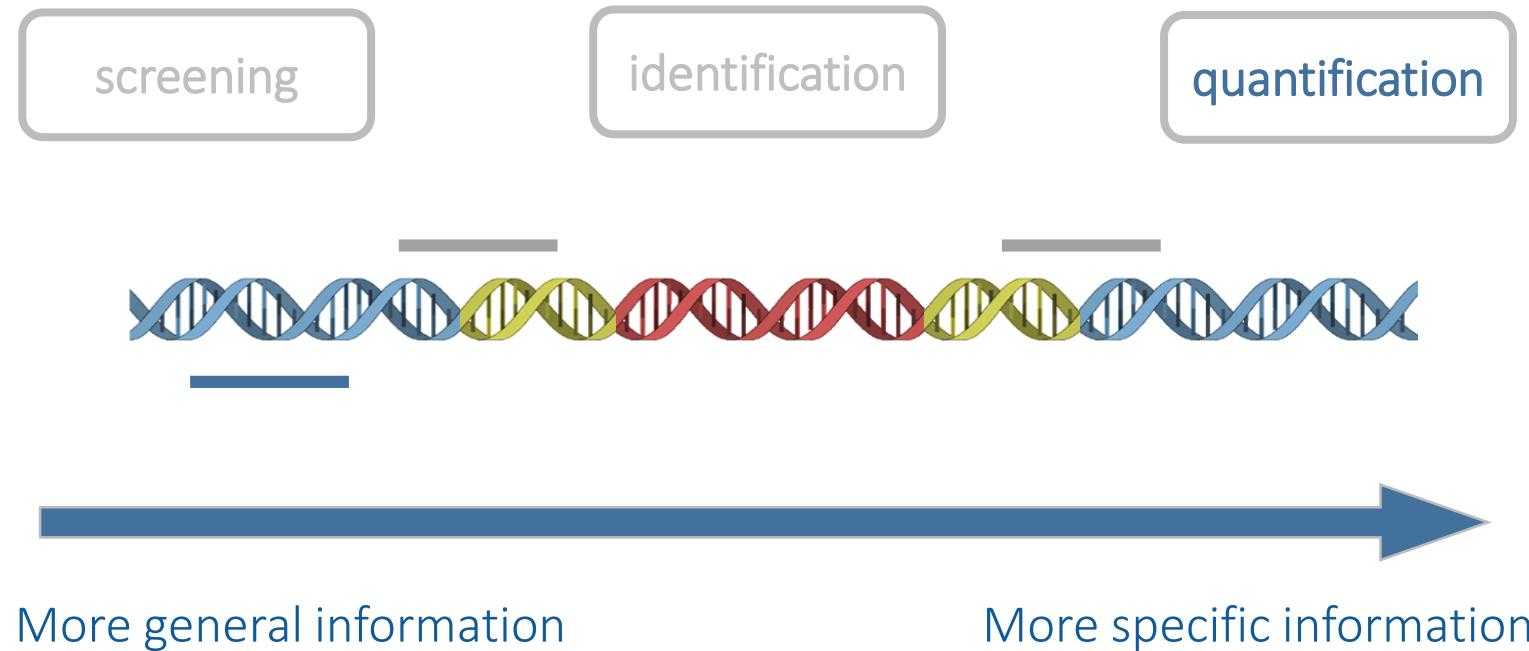
## Three partite PCR-based testing scheme - screening



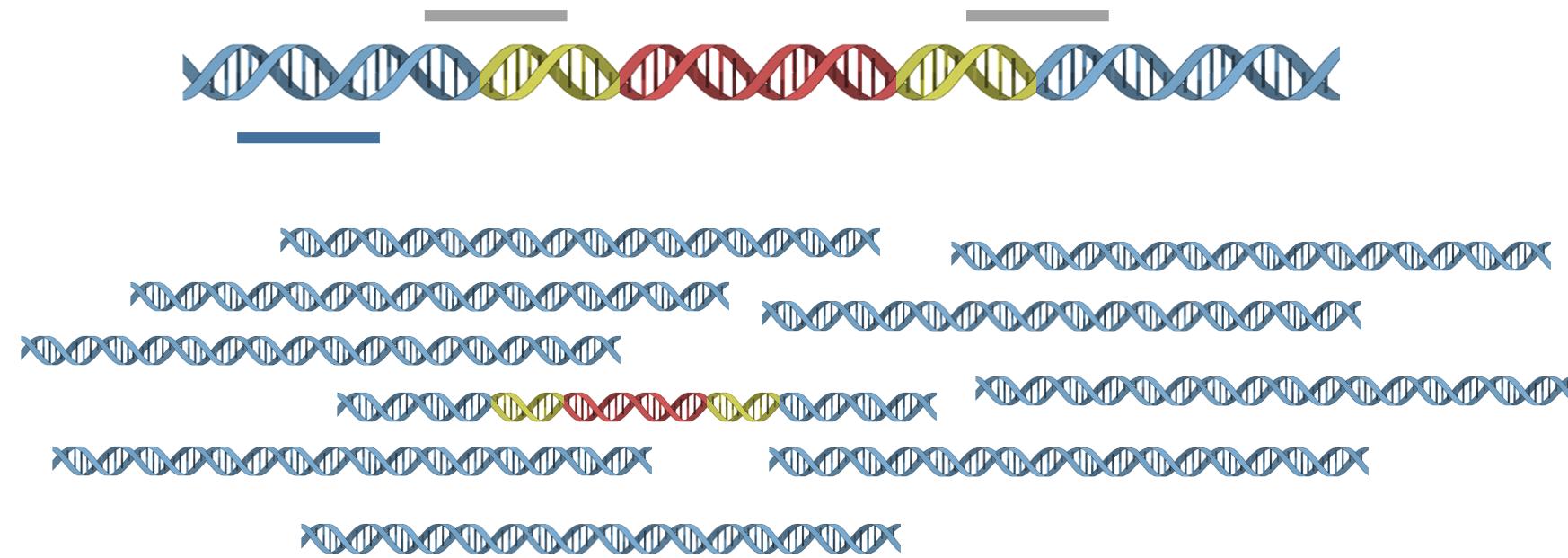
## Three partite PCR-based testing scheme - identification



# Three partite PCR-based testing scheme - quantification



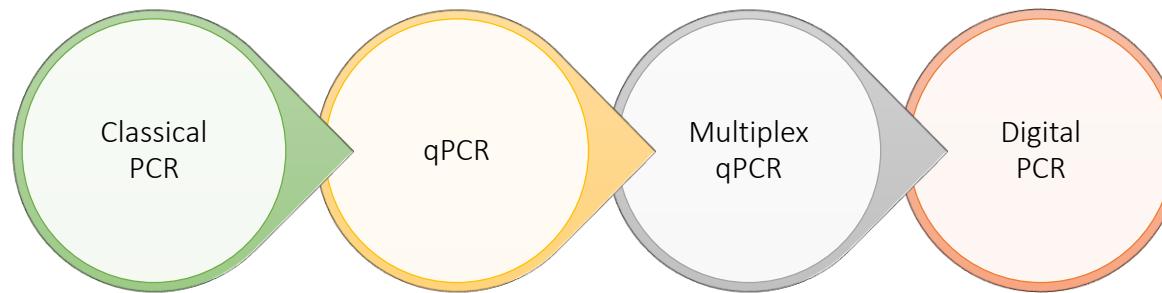
## Three partite PCR-based testing scheme - quantification



1 GM + 9 non GM = 10% GMO

# Challenges in GMO testing

- Unknown GMOs
- Unauthorized GMOs
- **Increasing number of GMOs**
- **GMOs without common elements**
- **GMOs in challenging matrices**



# Increasing number of GMOs

Screening for common elements and decision support after the screening

## 5 common elements in pentaplex:

P35S	promoter region of the cauliflower mosaic virus
Tnos	nopaline synthase gene terminator from <i>Agrobacterium tumefaciens</i>
bar	basta resistance gene from <i>Streptomyces hygroscopicus</i>
pat	phosphinothricine acetyltransferase gene from <i>Streptomyces viridochromogenes</i>
ctp-cp4-epsps	construct containing a 6-enolpyruvylshikimate-3-phosphate-synthase gene from <i>Agrobacterium tumefaciens</i> strain cp4 with an upstream sequence of the ctp2 chloroplast transit peptide from <i>Arabidopsis thaliana</i>

**GMOseek**

**GMOval project**

Real-time PCR screening methods



Select species	Screening assays	GMOs
<input checked="" type="checkbox"/> soybean	<input type="checkbox"/> CTP2-CP4-EPSPS	A2704-12 (ACS-GM005-3)
<input type="checkbox"/> rice	<input checked="" type="checkbox"/> tNOS	A5547-127 (ACS-GM006-4)
<input type="checkbox"/> linseed	<input checked="" type="checkbox"/> P35S	MON40-3-2 (MON-04032-6)
<input type="checkbox"/> bacteria	<input type="checkbox"/> bar	MON89788 (MON-89788-1)
<input type="checkbox"/> papaya	<input type="checkbox"/> pat	DP356043-5 (DP-356043-5)
<input type="checkbox"/> virus		DP305423-1 (DP-305423-1) LLP
<input checked="" type="checkbox"/> maize		CV127 (BPS-CV-127-9) LLP
<input type="checkbox"/> sugar beet		MON87701 (MON-87701-2)
<input type="checkbox"/> cotton		MON87769 (MON-87769-7) LLP
<input type="checkbox"/> oilseed rape		68416 (DAS-68416-4) LLP
<input type="checkbox"/> potato		MON87708 (MON-87708-9) LLP
		MON87705 (MON-87705-6) LLP
		FG72 (MST-FG072-2) LLP
		3272 (SYN-E3272-5) LLP
		Bt11 (SYN-BT011-1)
		DAS59122 (DAS-59122-7)
		GA21 (MON-00021-9)
		MIR604 (SYN-IR604-5)
		MON810 (MON-00810-6)
		MON863 (MON-00863-5)
		NK603 (MON-00603-6)
		T25 (ACS-ZM003-2)
		DAS1507 (DAS-01507-1)
		MON89034 (MON-89034-3)
		MON88017 (MON-88017-3)
		MIR162 (SYN-IR162-4)
		40278 (DAS-40278-9) LLP
		MON87460 (MON-87460-4) LLP

Select all      Unselect all

Results of testing feed (containing soybean and maize) with pentaplex:

P-35S - present

Tnos - present

bar – not present

pat - not present

ctp-cp4-epsps - not present

In the next step presence of 8 soybean lines and 9 maize lines should be tested (marked in red)

# Increasing number of GMOs

Screening for common elements and decision support after the screening

JOURNAL OF  
**AGRICULTURAL AND  
FOOD CHEMISTRY**

Article

[pubs.acs.org/JAFC](https://pubs.acs.org/JAFC)

## Development and Validation of Duplex, Triplex, and Pentaplex Real-Time PCR Screening Assays for the Detection of Genetically Modified Organisms in Food and Feed

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<sup>‡</sup>Walloon Agricultural Research Centre (CRA-W), Chaussée de Namur 24, B-5030 Gembloux, Belgium

<sup>§</sup>Department of Biotechnology and Systems Biology, National Institute of Biology (NIB), Vecna pot 111, SI-1000 Ljubljana, Slovenia

<sup>⊥</sup>Federal Office of Consumer Protection and Food Safety (BVL), Mauerstrasse 39-42, 10117 Berlin, Germany



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# GMOs without common elements

With screening for selected common elements it is not possible to detect all in EU authorized GMOs

Event (Unique identifier)	Species	CTP2-CP4- EPSPS	Tnos	P35S	bar	pat
A2704-12 (ACS-GMØØ5-3)	soybean	0	0	1	0	1
A5547-127 (ACS-GMØØ6-4)	soybean	0	0	1	0	1
MON40-3-2 (MON-Ø4Ø32-6)	soybean	0	1	1	0	0
MON89788 (MON-89788-1)	soybean	1	0	0	0	0
DP356043-5 (DP-356Ø43-5)	soybean	0	0	1	0	0
DP305423-1 (DP-3Ø5423-1)	soybean	0	0	0	0	0
CV127 (BPS-CV-127-9)	soybean	0	0	0	0	0
MON87701 (MON-877Ø1-2)	soybean	0	0	0	0	0
MON87769 (MON-87769-7)	soybean	0	0	0	0	0
68416 (DAS-68416-4)	soybean	0	0	0	0	1
MON87708 (MON-87708-9)	soybean	0	0	0	0	0
MON87705 (MON-877Ø5-6)	soybean	1	0	0	0	0
FG72 (MST-FGØ72-2)	soybean	0	1	0	0	0
DAS44406 (DAS-44406-6)	soybean	0	0	0	0	1
DAS81419 (DAS-81419-2)	soybean	0	0	0	0	1
SYHTOH2 (SYN-ØØØH2-5)	soybean	0	1	1	0	1
MON87751 (MON-87751-7)	soybean	0	0	0	0	0
GMB151 (BCS-GM151-6)	soybean	0	0	1	0	0

Event (Unique identifier)	Species	CTP2-CP4- EPSPS	Tnos	P35S	bar	pat
A2704-12 (ACS-GMØØ5-3)	soybean	0	0	1	0	1
A5547-127 (ACS-GMØØ6-4)	soybean	0	0	1	0	1
MON40-3-2 (MON-Ø4Ø32-6)	soybean	0	1	1	0	0
MON89788 (MON-89788-1)	soybean	1	0	0	0	0
DP356043-5 (DP-3Ø543-5)	soybean	0	0	1	0	0
DP305423-1 (DP-3Ø5423-1)	soybean	0	0	0	0	0
CV127 (BPS-CV-127-9)	soybean	0	0	0	0	0
MON87701 (MON-877Ø1-2)	soybean	0	0	0	0	0
MON87769 (MON-87769-7)	soybean	0	0	0	0	0
68416 (DAS-68416-4)	soybean	0	0	0	0	1
MON87708 (MON-87708-9)	soybean	0	0	0	0	0
MON87705 (MON-877Ø5-6)	soybean	1	0	0	0	0
FG72 (MST-FGØ72-2)	soybean	0	1	0	0	0
DAS44406 (DAS-44406-6)	soybean	0	0	0	0	1
DAS81419 (DAS-81419-2)	soybean	0	0	0	0	1
SYHTOH2 (SYN-ØØØH2-5)	soybean	0	1	1	0	1
MON87751 (MON-87751-7)	soybean	0	0	0	0	0
GMB151 (BCS-GM151-6)	soybean	0	0	1	0	0

Event (Unique identifier)	Species	CTP2-CP4- EPSPS	Tnos	P35S	bar	pat
A2704-12 (ACS-GMØØ5-3)	soybean	0	0	1	0	1
A5547-127 (ACS-GMØØ6-4)	soybean	0	0	1	0	1
MON40-3-2 (MON-Ø4Ø32-6)	soybean	0	1	1	0	0
MON89788 (MON-89788-1)	soybean	1	0	0	0	0
DP356043-5 (DP-3Ø543-5)	soybean	0	0	1	0	0
DP305423-1 (DP-3Ø5423-1)	soybean	0	0	0	0	0
CV127 (BPS-CV-127-9)	soybean	0	0	0	0	0
MON87701 (MON-877Ø1-2)	soybean	0	0	0	0	0
MON87769 (MON-87769-7)	soybean	0	0	0	0	0
68416 (DAS-68416-4)	soybean	0	0	0	0	1
MON87708 (MON-87708-9)	soybean	0	0	0	0	0
MON87705 (MON-877Ø5-6)	soybean	1	0	0	0	0
FG72 (MST-FGØ72-2)	soybean	0	1	0	0	0
DAS44406 (DAS-44406-6)	soybean	0	0	0	0	1
DAS81419 (DAS-81419-2)	soybean	0	0	0	0	1
SYHTOH2 (SYN-ØØØH2-5)	soybean	0	1	1	0	1
MON87751 (MON-87751-7)	soybean	0	0	0	0	0
GMB151 (BCS-GM151-6)	soybean	0	0	1	0	0

Instead of screening for common elements, we are screening for presence of all in EU authorized lines:

The same set up of the qPCR for all runs

Plates (384 wells) for qPCR are partly prepared in advance – primers and probes are distributed on the plate and stored <-15°C

Samples and controls are mixed with the mastermix for qPCR and added to the plate



# GMOs in challenging matrices



Contents lists available at [ScienceDirect](#)

Food Chemistry

journal homepage: [www.elsevier.com/locate/foodchem](http://www.elsevier.com/locate/foodchem)

Digital PCR as an effective tool for GMO quantification in complex matrices

Alexandra Bogožalec Košir\*, Tina Demšar, Dejan Štebih, Jana Žel, Mojca Milavec

*Department of Biotechnology and Systems Biology, National Institute of Biology, Večna pot 111, SI-1000 Ljubljana, Slovenia*

# GMOs in challenging matrices



VS.



© Bio-Rad

VS.



Use of dPCR for quantification of GM soybean line in complex samples, containing PCR inhibitors.

## GMOs in challenging matrices

Feed sample containing GM soybean – MON40-3-2 (Roundup Ready)

qPCR – sample exhibits great inhibition for both event specific sequence (MON40-3-2) and reference gene/endogene (Le1) up to 100 × dilution, sample cannot be quantified → pLOQ = 49 %

Methods (validated) were transferred to dPCR

## GMOs in challenging matrices

Feed sample containing GM soybean – MON40-3-2 (Roundup Ready)

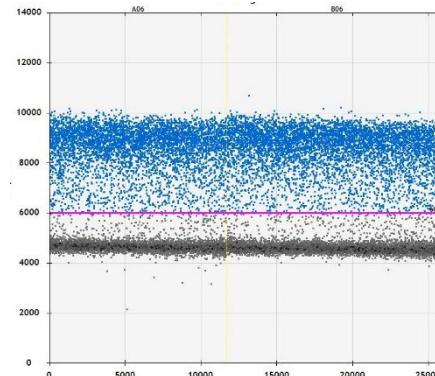
qPCR – sample exhibits great inhibition for both event specific sequence (MON40-3-2) and reference gene/endogene (Le1) up to 100 × dilution, sample cannot be quantified → pLOQ = 49 %

Methods (validated) were transferred to dPCR

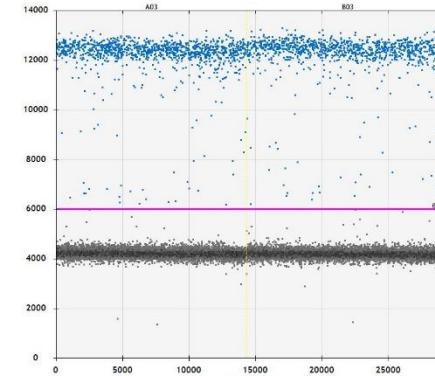
ddPCR	cdPCR	Bias
<b>35.45</b>	<b>31.07</b>	<b>14.09 %</b>

# dPCR is not completely „immune“ to inhibitors

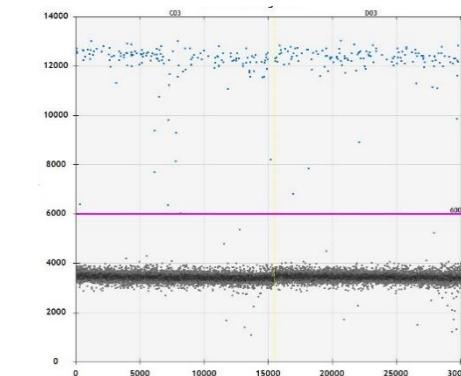
1× dilution – 14000 cp/µL



10× dilution – 2300 cp/µL

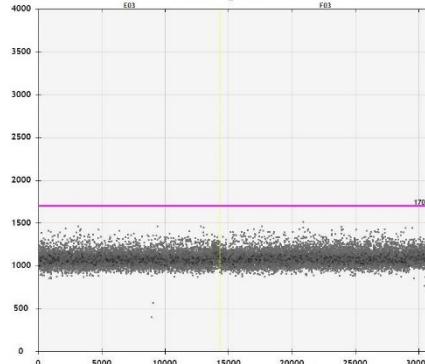


100× dilution – 260 cp/µL

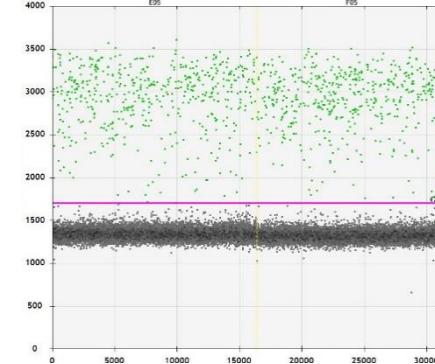


Reference gene

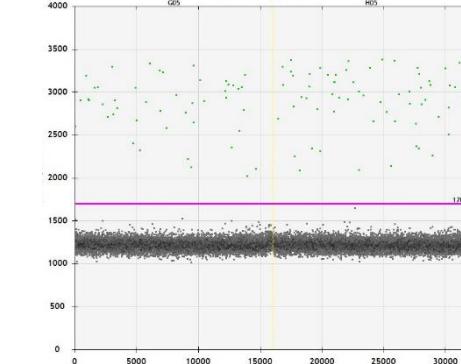
1× dilution – 0 cp/µL



10× dilution – 810 cp/µL



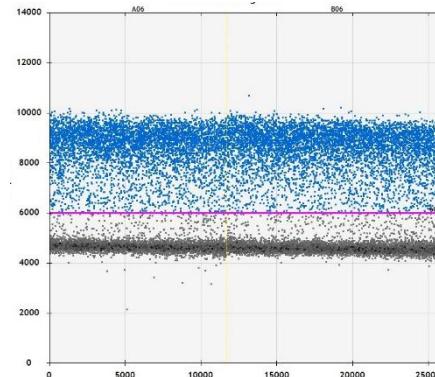
100× dilution – 90 cp/µL



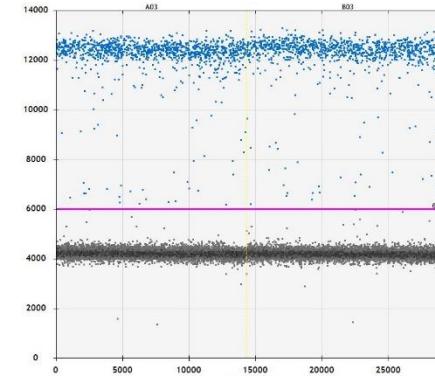
GMO

# dPCR is not completely „immune“ to inhibitors

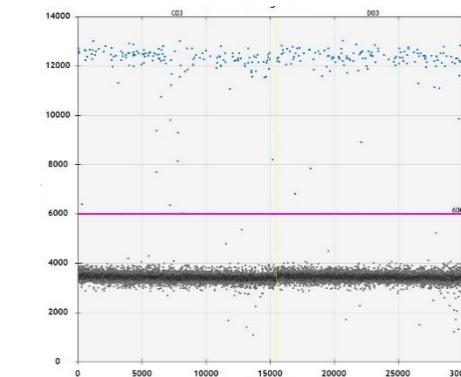
1× dilution – 14000 cp/µL



10× dilution – 2300 cp/µL

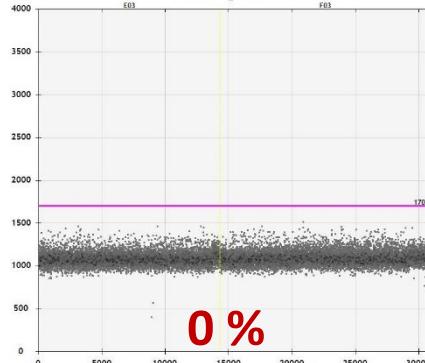


100× dilution – 260 cp/µL

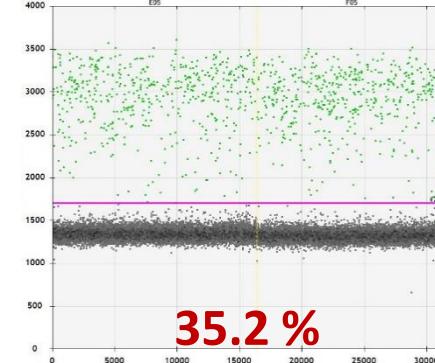


Reference gene

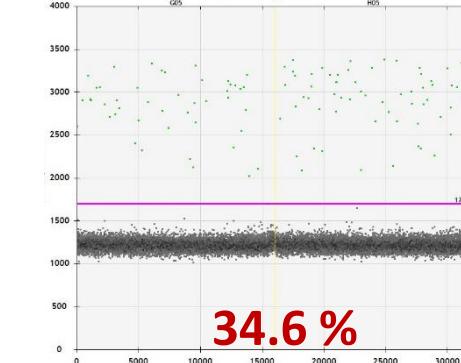
1× dilution – 0 cp/µL



10× dilution – 810 cp/µL



100× dilution – 90 cp/µL



GMO

# Accreditation

2 simplex assays

Soybean endogene *Le1*

MON40-3-2 or Roundup Ready soybean

A duplex assay for quantification of MON40-3-2



Reg. šL / Ref. No.: 3150-0064/10-0006

Datum izdaje / Issued on: 10. oktober 2016

Zamenjuje izdajo z dne / Replaces Annex dated: 12. marec 2015

Voljavnost akreditacije je mogoča preveriti na spletni strani SA, www.slo-akreditacija.si.  
Information on current accreditation status is available at the SA website, www.slo-akreditacija.si.

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**LP-028**

## PRILOGA K AKREDITACIJSKI LISTINI Annex to the accreditation certificate

**LP-028**

### 1 AKREDITIRANI ORGAN / Accredited body

Nacionalni inštitut za biologijo  
Večna pot 111, 1000 Ljubljana

Kopija pristojne objave na spletnem mestu. / Copy of attachment for web publishing.

### 2 STANDARD

SIST EN ISO/IEC 17025:2005

### 3 OBSEG AKREDITACIJE / Scope of accreditation

V okviru te akreditacijske listine Slovenska akreditacija priznava akreditiranemu organu usposobljenost za opravljanje naslednjih dejavnosti: / SA hereby acknowledges the accredited body as being competent for performing the following activities:

#### 3.1 Skrajšan opis obsega akreditacije / A short description of the scope

Področja preskušanja glede na vrsto preskušanja / Testing fields with reference to the type of test:

- biologija, biokemijski (določanje GSO) / biology, biochemical (GMO detection)
- mikrobiologija (molekularne metode) / microbiology (molecular methods)

Področja preskušanja glede na vrsto preskušanca / Testing fields with reference to the type of test item:

- živila / foodstuffs
- kmetijski proizvodi / agricultural products
- biološki vzorci / biological samples



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# Plant Pathogens

Fire blight (*Erwinia amylovora*)



Potato brown rot (*Ralstonia solanacearum*)



Anal Bioanal Chem  
DOI 10.1007/s00216-014-8084-1

PAPER IN FOREFRONT

## Optimising droplet digital PCR analysis approaches for detection and quantification of bacteria: a case study of fire blight and potato brown rot

Tanja Drešo · Manca Pirc · Živa Ramšak · Jernej Pavšič ·  
Mojca Milavec · Jana Žel · Kristina Gruden

Received: 19 June 2014 / Revised: 28 July 2014 / Accepted: 29 July 2014  
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# Human Pathogens

## Human cytomegalovirus (HCMV)

Anal Bioanal Chem (2016) 408:107–121  
DOI 10.1007/s00216-015-9107-2



PAPER IN FOREFRONT

### Assessment of the real-time PCR and different digital PCR platforms for DNA quantification

Jernej Pavšič<sup>1,2</sup> · Jana Žel<sup>1</sup> · Mojca Milavec<sup>1</sup>

Anal Bioanal Chem (2016) 408:67–75  
DOI 10.1007/s00216-015-9109-0



RAPID COMMUNICATION

### Digital PCR for direct quantification of viruses without DNA extraction

Jernej Pavšič<sup>1,2</sup> · Jana Žel<sup>1</sup> · Mojca Milavec<sup>1</sup>



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Thank you for your attention

