

"Enhancing Collaboration between the CEE and Central Asia's Centres of Excellence to Address the Key Drivers of Biodiversity Loss and Maintain Human, Crop and Livestock Health"

Workshop on Laboratory Identification of Species, Screening of Living Modified Organisms and Detection of Plant Pathogens

February 12-16, 2024



Convention on
Biological Diversity



ENVIRONMENTAL DNA (eDNA): A REVOLUTIONARY APPROACH TO STUDY BIODIVERSITY OF ANY ENVIRONMENT



Dr. Bojana Mićić,

Department of Biochemistry

Institute for Biological Research „Siniša Stanković“

National Institute of the Republic of Serbia

University of Belgrade

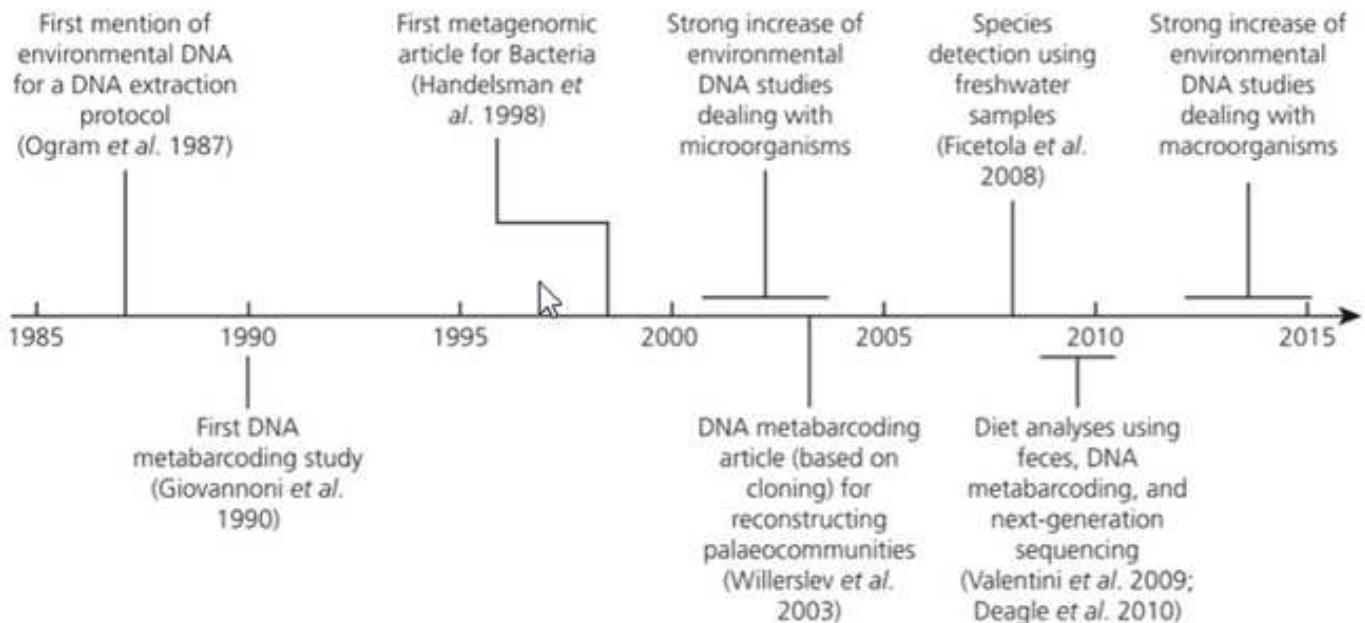
Institute of Genetics and Cytology of the National Academy of Sciences of Belarus (IGC)
Minsk, Republic of Belarus

Environmental DNA (eDNA)

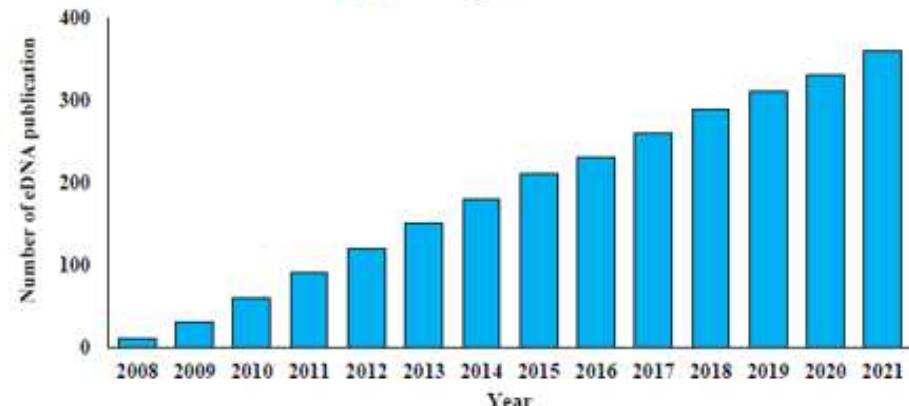
- **Nuclear or mitochondrial DNA discharged to the environment** by the organisms (faeces, urine, mucus, epidermal cells, gamets, corpses...)
- Usually short, degraded fragments shorter than 500 bp
- Intracellular and extracellular DNA
- The rate of degradation varies depending on different environmental and biological factors



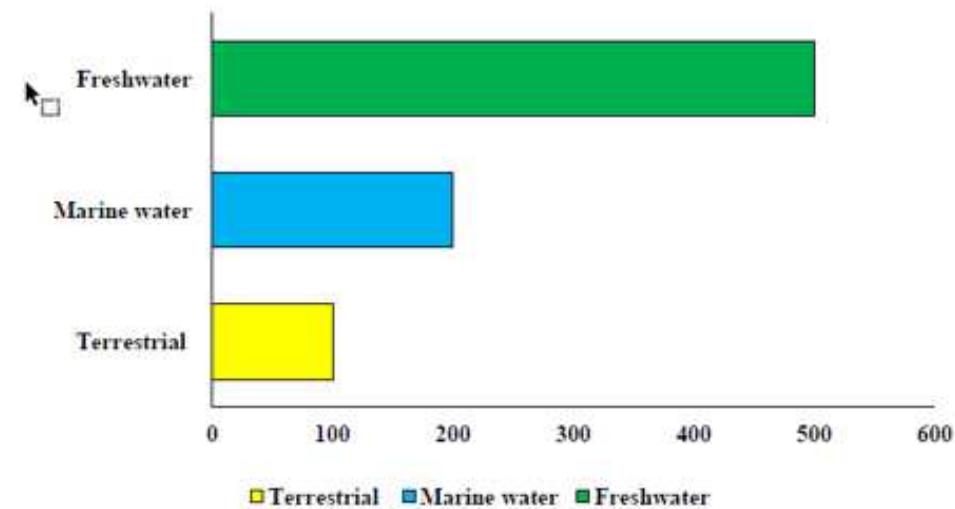
A brief history of eDNA



■ Number of publications

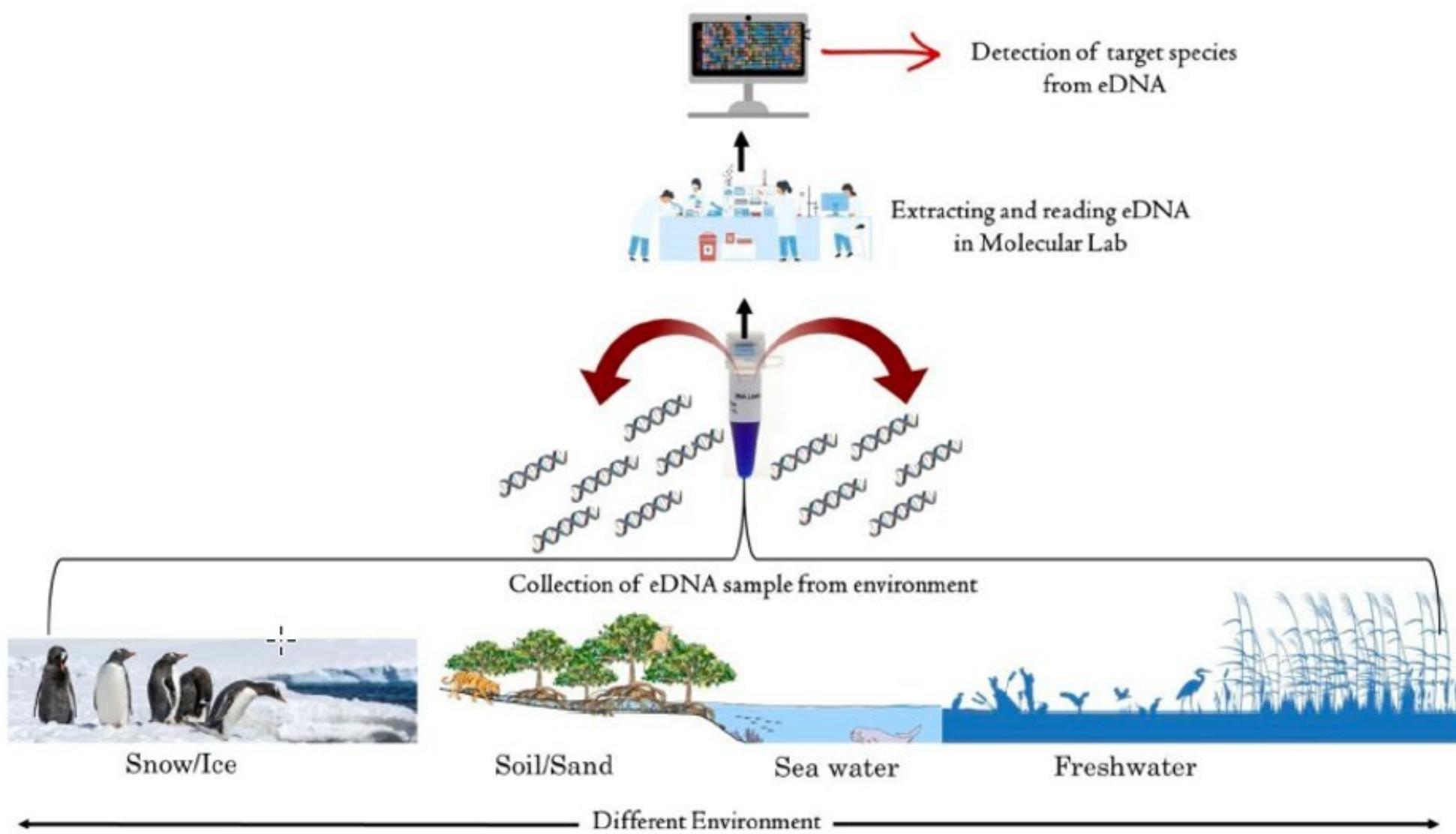


Number of studies using environmental DNA (eDNA) increasing day by day between 2008 and 2021.



Number of studies using environmental DNA (eDNA) in different environments

eDNA workflow



eDNA workflow

- **eDNA extraction**
 - Water, air, soil, sediment...
 - Grab or composite sampling?
 - Sterilization of the equipment to avoid contamination!
- **eDNA capture**
 - Filtration (glass fiber, nitrocellulose, PES)
 - Precipitation (ethanol/isopropanol + sodium acetate)
 - Centrifugation
- **eDNA extraction**
 - Commercial extraction kits
- **Analysis**

Step 1: Choose the best protocol, depending on your conditions



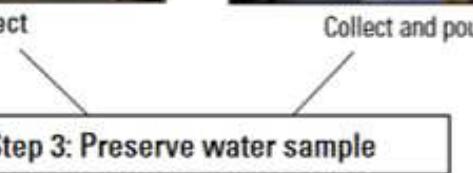
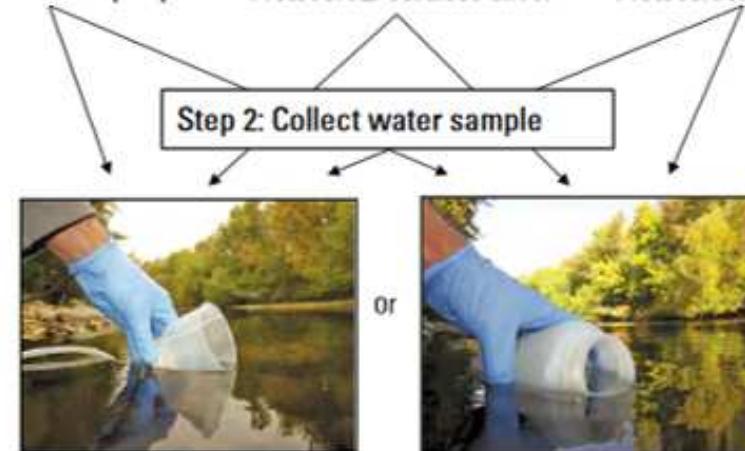
Protocol #1: Hand pump



Protocol #2: Cordless driver



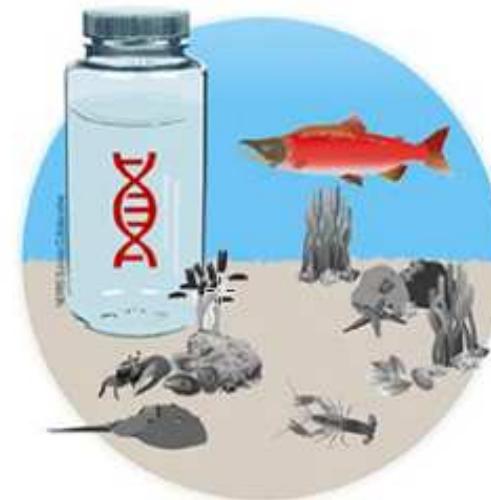
Protocol #3: 120-V pump



Ethanol-filled vial

Analysis approaches

- **eDNA barcoding**
 - Target species analysis
 - PCR with species-specific primers
 - Conventional PCR methods
- **eDNA metabarcoding**
 - To study multiple species and community dynamics
 - High-throughput sequencing methods
 - Selection of a target region



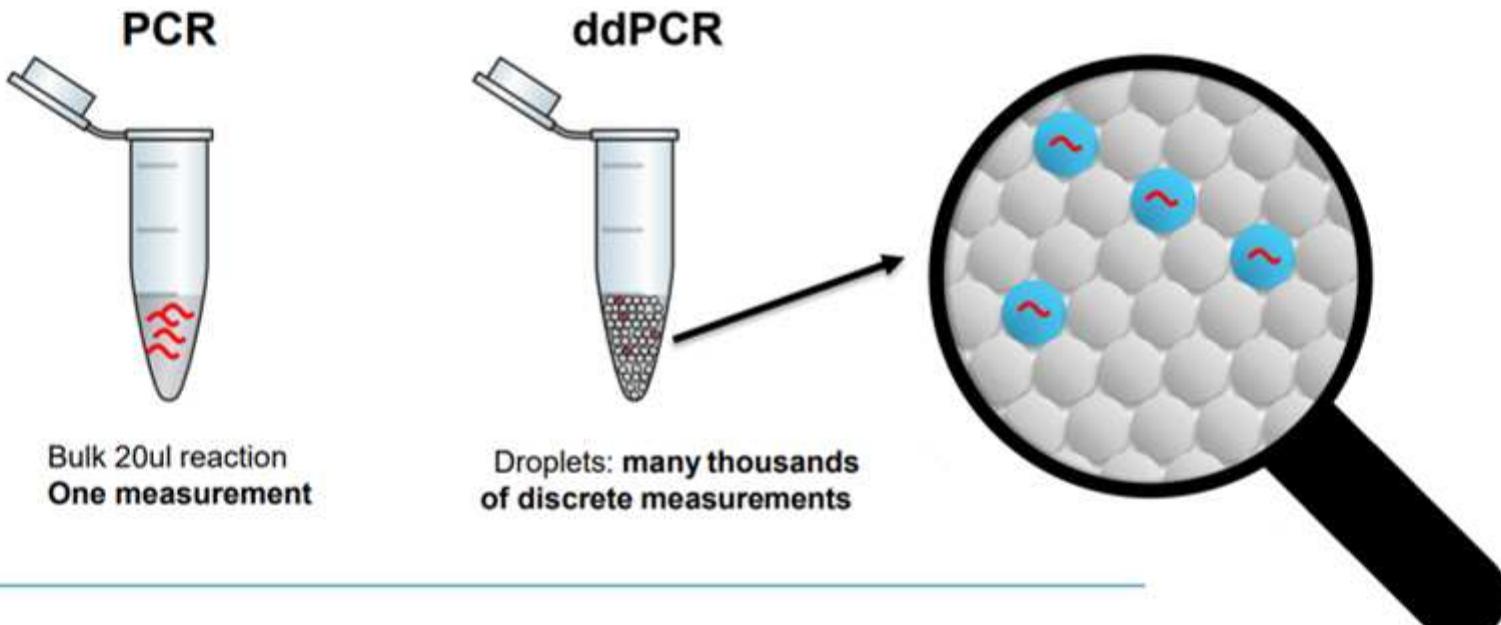
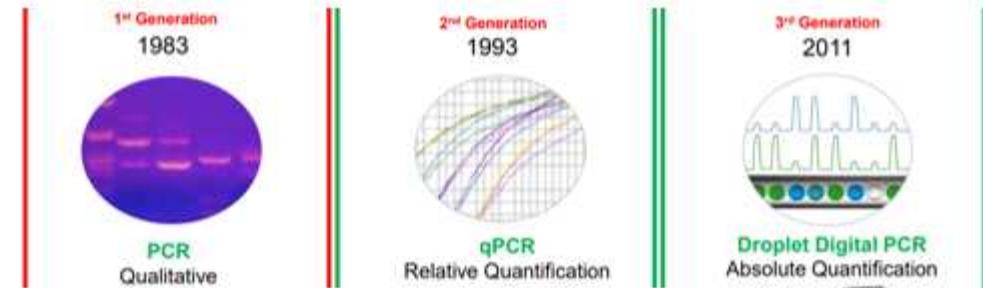
Single Species PCR



Metabarcoding

Droplet digital PCR (ddPCR)

- 3rd generation PCR (2011)
- Absolute quantification

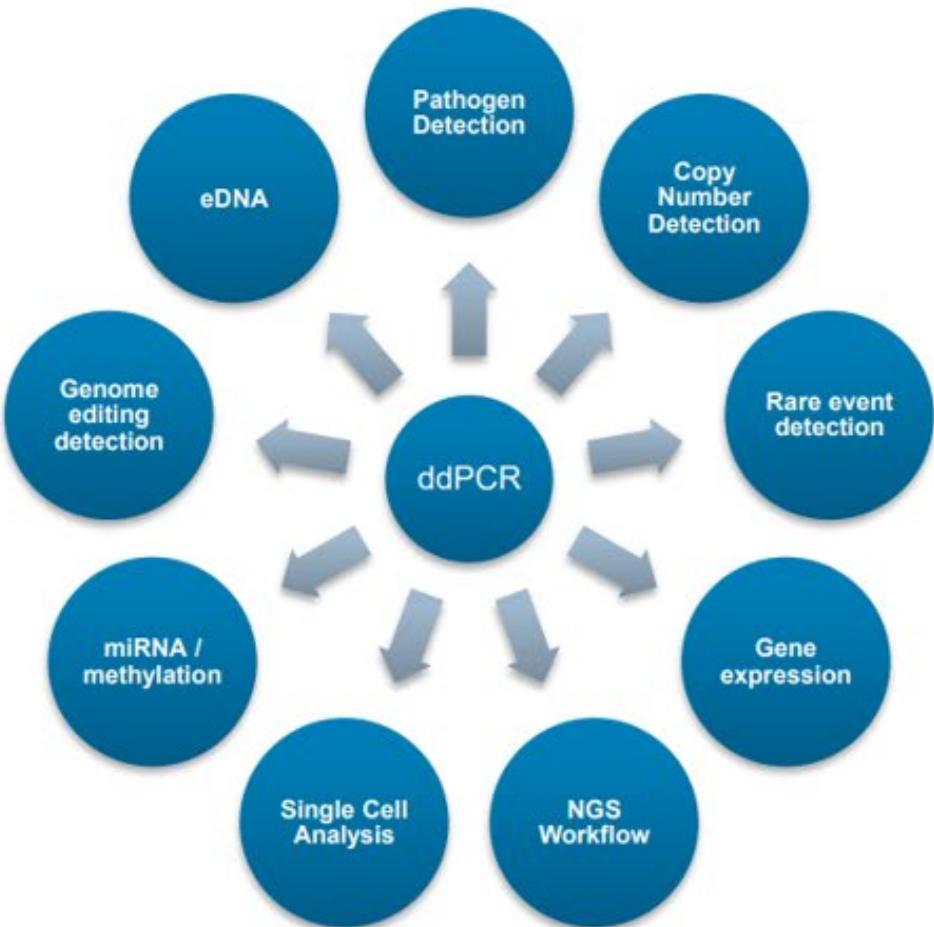


One partition → One micro amplification → One data analysis

Key advantages of ddPCR

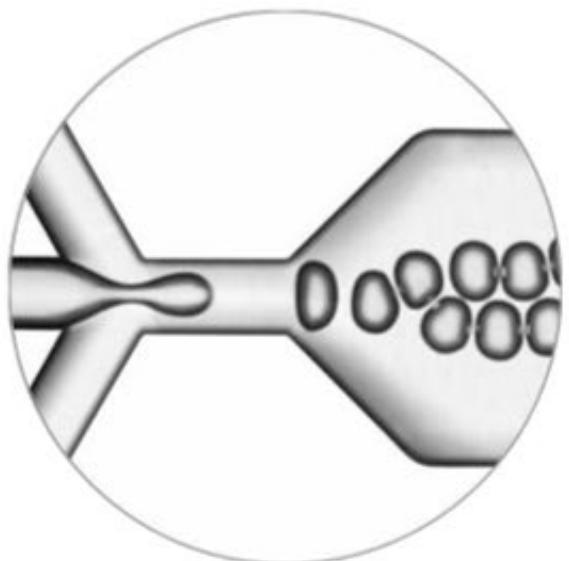
- Absolute quantification (no standard curve required)
- Endpoint method
- Less sensitive to PCR inhibition
- Reduced intra- and interlaboratory variation
- Quantitate low input concentration of DNA target
- Quantitate a rare target in large backgrounds

Possible uses of ddPCR

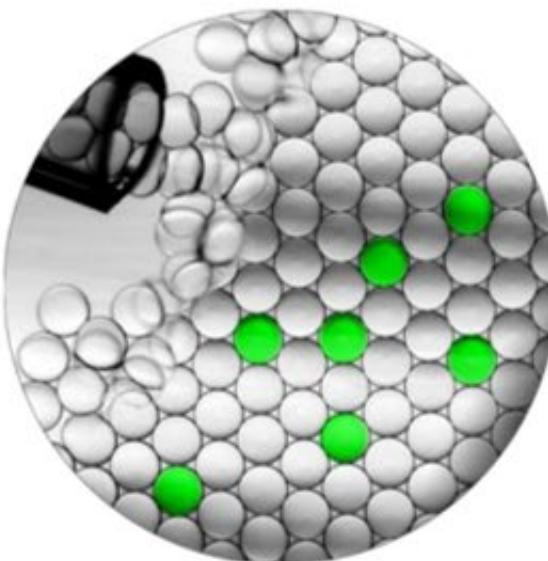


ddPCR workflow

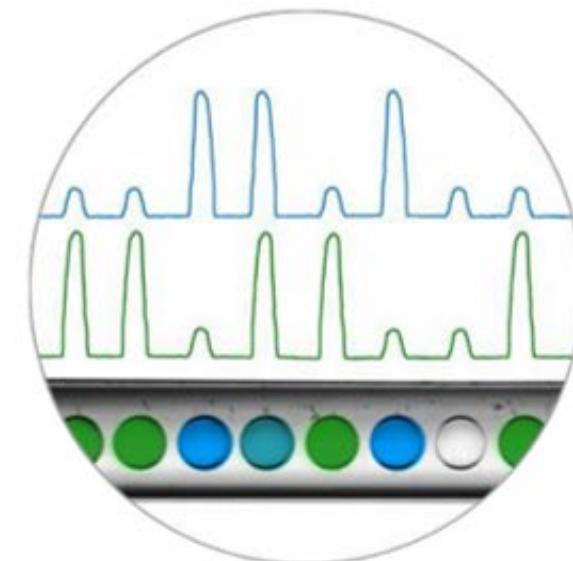
Partition sample
into Droplets



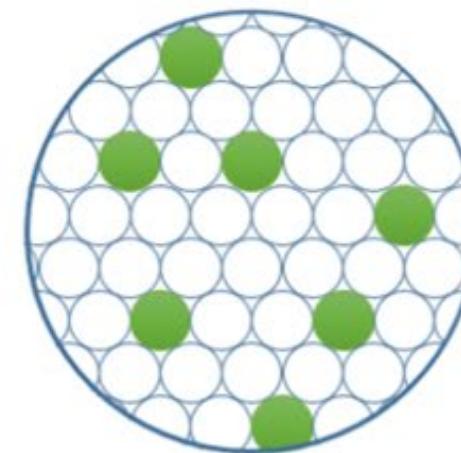
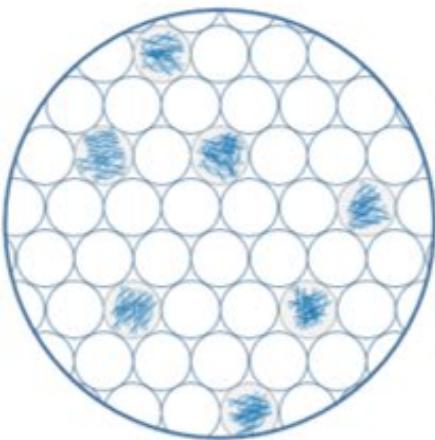
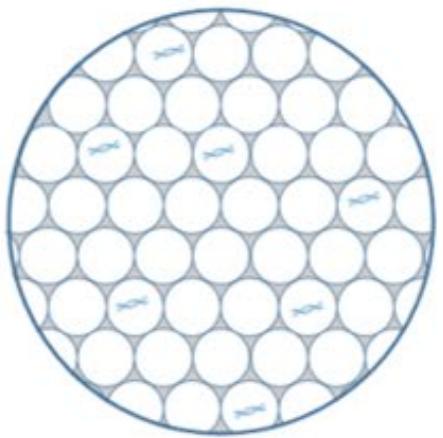
Cycle Droplets



Read
Droplets

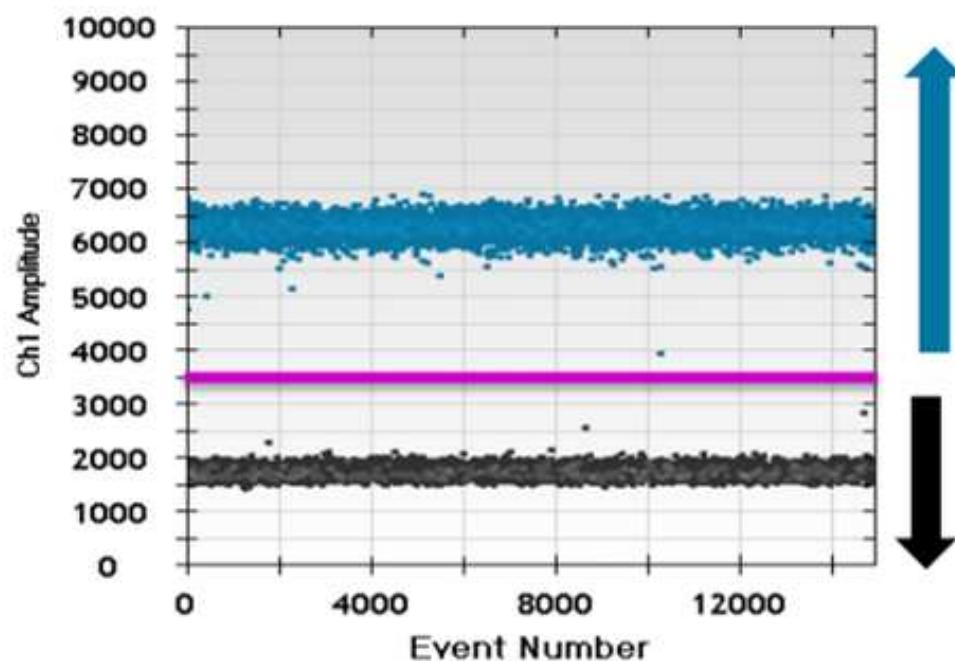


ddPCR workflow



Interpretation of the results

- Positive droplets contain at least one copy of target DNA or cDNA
- Increased fluorescence in positive vs. negative droplets
- Software measures the number of positive and negative droplets per fluorophore per sample



Each positive counted as 1

Threshold

Each negative counted as 0

What we've been doing so far

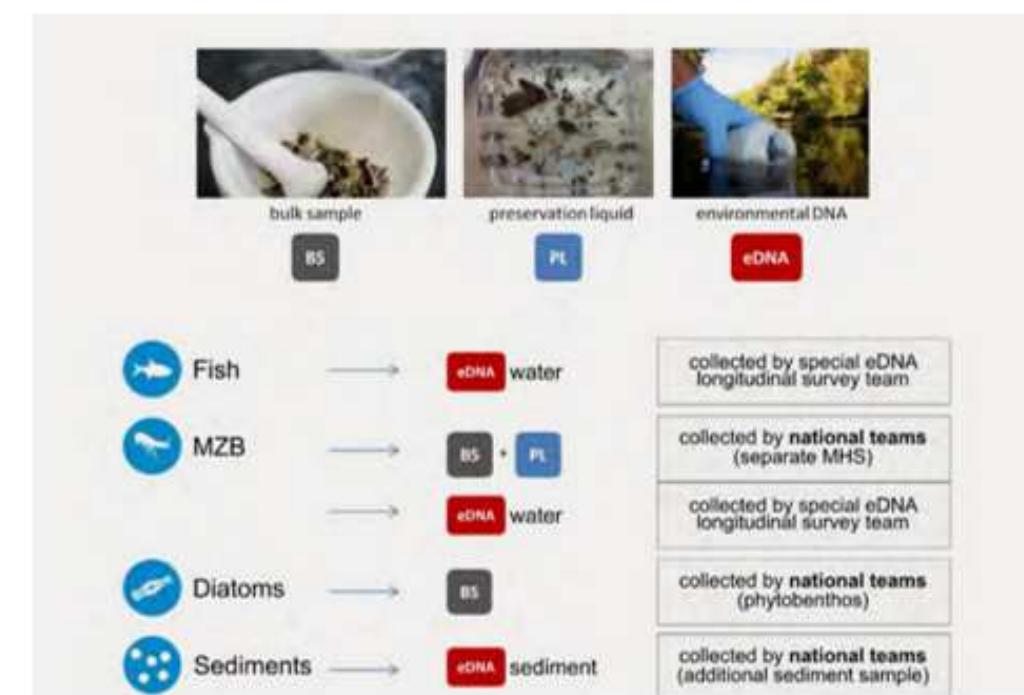
- The TransNational Monitoring Network is an important tool under the Danube River Protection Convention (1996)
- Aims to provide a well-balanced overall view of pollution and long-term trends in water quality and pollution loads in the Danube and its major tributaries
- The EU Water Framework Directive (WFD) requires that countries in the Danube River Basin periodically assess certain water characteristics in their territory.
- JDS1 (2001), JDS2 (2006), JDS3 (2013), **JDS4 (2019)**

JOINT
DANUBE
SURVEY 4



What we've been doing so far

- With JDS4, eDNA-based metabarcoding approach included for the first time in the extensive analytical program of the Joint Danube Survey
- Fish, macrozoobenthos, phytobenthos and sediment community
- 18S, COI, rbcL, 12S
- Promising potential of the eDNA metabarcoding approach recognised
- Coherent results when compared to traditional capture methods
- Significant increase in taxonomic resolution
- The detection of hard to observe species and developmental stages enabled
- Still can be improved and standardised, JDS5 is being planned at the moment...



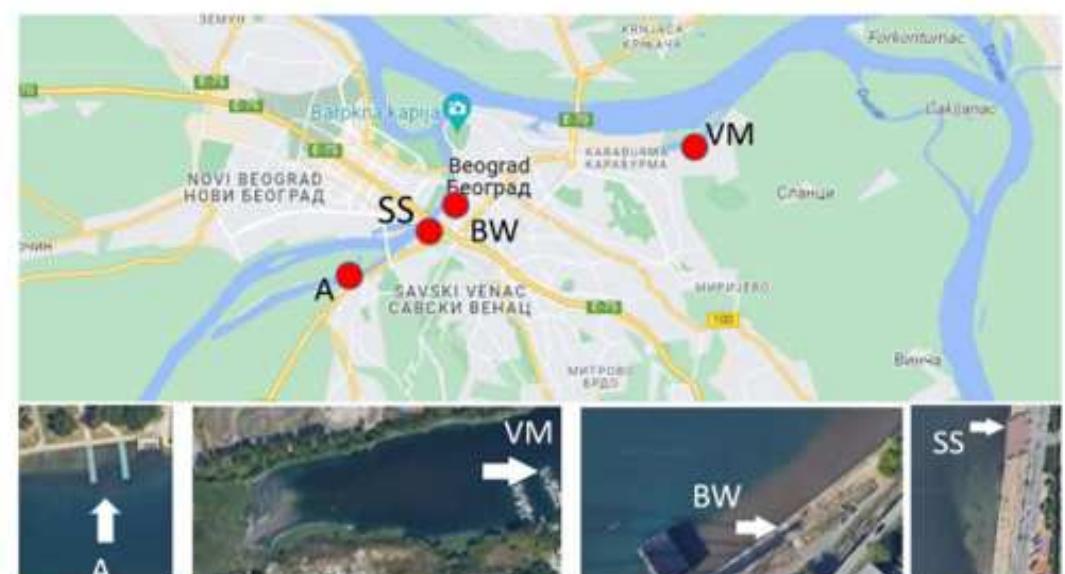
What we've been doing so far

- **Wastewater based epidemiology approach**
- Bilateral cooperation agreement with Belarus: IBISS + The Republican Research and Practical Center for Epidemiology and Microbiology (Minsk)
- Commercial application of ddPCR in wastewater-based epidemiology
- Development and implementation of common algorithm for monitoring SARS-CoV2 based on wastewater epidemiology
- Development of **early warning system** which, based on the presence of SARS-CoV-2 RNA in wastewater, would indicate the presence of infected persons in the population
- Wastewater, surface water or tissues of filtering-feeding sessile benthic animals as a sample?



What we've been doing so far

- **Wastewater based epidemiology approach: Belgrade**
- Samples: RNA extracts from wastewaters; RNA extracts from digestive gland of *Sinanodonta woodiana*
- RNeasy mini kit (Qiagen, Cat. No. 74104)
- PREvalence ddPCR SARS-CoV-2 Wastewater Quantification
- Multiplex: N2 and E regions





For further cooperation, please contact:
Dr. Stoimir Kolarević, senior research associate
Head of ddPCR facility at IBISS
stoimir.kolarevic@ibiss.bg.ac.rs



Thank you for your attention!
Спасибо за внимание!
Dr. Bojana Mićić, research assistant
bojana.micic@ibiss.bg.ac.rs

Suggested resources to start with ☺

<https://ednaresources.science/>

<https://www.bio-rad.com/en-rs/life-science/droplet-digital-pcr>

<https://doi.org/10.1007/s10531-020-01980-0>

<https://doi.org/10.1016/j.jnc.2022.126325>